

# FavorPrep™ Viral DNA/RNA Kit

For isolation of viral DNA/ RNA from cell-free fluids such as, serum, plasma, body fluids and cell-cultured supernatant, via transport medium of swabs

## For Research Use Only

## **Kit Contents:**

Cat. No:	FAVNK 000-1 (4 preps_sample)	FAVNK 001 (50 preps)
VNE Buffer ■	1.8 ml x 2	35 ml
Carrier RNA ■	0.04 mg	0.4 mg
Wash Buffer 1 □ !!! (concentrate)	0.48 ml x 2	12 ml
Wash Buffer 2 □ (concentrate)	1.5 ml	20 ml
RNase-free Water	0.5 ml	6 ml
VNE Column	4 pcs	50 pcs
Collection Tube	8 pcs	100 pcs
Elution Tube	4 pcs	50 pcs
User Manual	1	1

■ □ Adding Ethanol to the concentrate Wash Buffer. see Working Buffer Preparation.

!!! Note: the ethanol percentage of Wash Buffer 1 has changed (Version. Jan 21). see Working Buffer Preparation

Cat. No:       FAVNK 001-1 (100 preps)       FAVNK 001-2 (300 preps)         VNE Buffer ■       70 ml       200 ml         Carrier RNA ■       0.8 mg       2.2 mg         Wash Buffer 1 □ !!! (concentrate)       24 ml       72 ml         Wash Buffer 2 □ (concentrate)       20 ml x 2       50 ml x 2         RNase-free Water       12 ml       20 ml         VNE Column       100 pcs       300 pcs         Collection Tube       200 pcs       600 pcs         Elution Tube       100 pcs       300 pcs			
Carrier RNA ■       0.8 mg       2.2 mg         Wash Buffer 1 □ !!! (concentrate)       24 ml       72 ml         Wash Buffer 2 □ (concentrate)       20 ml x 2 (concentrate)       50 ml x 2         RNase-free Water       12 ml       20 ml         VNE Column       100 pcs       300 pcs         Collection Tube       200 pcs       600 pcs	Cat. No:		
Wash Buffer 1 □ !!! (concentrate)       24 ml       72 ml         Wash Buffer 2 □ (concentrate)       20 ml x 2       50 ml x 2         RNase-free Water       12 ml       20 ml         VNE Column       100 pcs       300 pcs         Collection Tube       200 pcs       600 pcs	VNE Buffer ■	70 ml	200 ml
(concentrate)       20 ml x 2       50 ml x 2         (concentrate)       20 ml x 2       20 ml x 2         RNase-free Water       12 ml       20 ml         VNE Column       100 pcs       300 pcs         Collection Tube       200 pcs       600 pcs	Carrier RNA ■	0.8 mg	2.2 mg
(concentrate)  RNase-free Water 12 ml 20 ml  VNE Column 100 pcs 300 pcs  Collection Tube 200 pcs 600 pcs		24 ml	72 ml
VNE Column 100 pcs 300 pcs Collection Tube 200 pcs 600 pcs		20 ml x 2	50 ml x 2
Collection Tube 200 pcs 600 pcs	RNase-free Water	12 ml	20 ml
	VNE Column	100 pcs	300 pcs
Elution Tube 100 pcs 300 pcs	Collection Tube	200 pcs	600 pcs
	Elution Tube	100 pcs	300 pcs
User Manual 1 1	User Manual	1	1

■ □ Adding Ethanol to the concentrate Wash Buffer. see Working Buffer Preparation.

!!! Note: the ethanol percentage of Wash Buffer 1 has changed (Version. Jan 21). see Working Buffer Preparation

## Storage

- Kit components (except Carrier RNA) should be stored at room temperatures between 15 - 25°C.
- 2. Carrier RNA should be stored at -20 °C upon receipt.
- 3. VNE Buffer should be stored at 4 °C after adding Carrier RNA.

## **Quality Control:**

The quality of FavorPrep™ Viral DNA/ RNA Kit is tested on a lot-to-lot basis according to ISO quality management system.

## **Description:**

FavorPrep™ Viral DNA/ RNA Kit is an excellent tool for extraction of highly pure viral nucleic acid from viral cell-free specimens such as, serum, plasma, body fluids and cell-cultured supernatant, via transport medium of swabs. It is exlcusively intended for this use by trained professionals.

## **Protocol overview:**

The extraction method is silica-based with a chaotropic salt technology presence. The procedure involves lysis of virus for optimization of a binding condition that promotes the efficient penetration of the viral nucleic acid into the silica membrane. Carefully carried out centrifugation /vacuum protocols, plus Wash Buffers 1&2 quarntee contaminants including salts, metabolites.

nucleases, and other components of body fluids are removed. Finally, highly pure Viral RNA is eluted from the silica membrane. The provided Carrier RNA in conjunction with the unique lysis buffer enhance the process of binding nucleic acid to the silica membrane. This produces a substrate cleaved by the nuclease contained in the sample mixture; thus, elevating the integrity and recovery of viral nucleic acid. When compared with other harmful and time-consuming methods, such as phenol/ chloroform extraction and ethanol

precipitation, the FavorPrep™ Viral DNA/RNA Kit makes the

extraction of high-purity viral nucleic acid reliable and efficient by

shortening the handling time by 20 minutes per preparation.

## **Specifications:**

Format/ Principle: spin column/ silica membrane/ chaotropic salt Sample size: 140 µl cell-free fluid such as serums, plasma, body fluids, and cell-cultured supernatants

Operation time: < 20 min Recovery rate: 80 ~ 90 %

Length of recovery nucleic acid: > 200 bp Column Binding capacity: 60 µg RNA /column

Elution volume: 40 ~ 50 µl

Column applicability: centrifugation and vacuum

## Materials and equipment provided by the user

## For centrifuge processing:

- 1. A microcentrifugator with a rotor for 1.5 or 2.0 ml microcentrifuge tubes
- The centrifuge force should reach ~18,000 x g.
- 2. Sterile pipets, pipet tips and centrifuge tubes (1.5 ml, 2.0 ml).
- 3. 100 % ethanol
- 4. Vortexer
- 5. Tube heater set to 60  $^{\circ}\text{C}$  and 70  $^{\circ}\text{C}$  for 1.5 or 2.0 ml microcentri fuge tubes

## For vacuum processing:

- 1. A micro-centrifugator with a rotor for 1.5~2.0 ml micro-centrifuge tubes
  - The centrifuge force should reach ~18,000 x g.
- 2. A vacuum manifold containing adaptors for DG Column.
  - The vacuum source should reach 6 inches Hg.
  - Make snsure that the the manifold adaptor is fitable to the DG column tip.
  - Units and values at same pressure (1 atm)

Office and values at same pressure (1 a	uii <i>)</i>
unit	value
atmosphere (atm)	1.000
millimeter of mercury (mmHg)	760.000
inches of mercury (inHg)	29.290
pascal (Pa)	101,325.000
kilopascal (KPa)	101.325
torr (torr)	760.000
pound per square inch (psi, 1bs/in²)	14.700

- 3. Sterile pipets, pipet tips and centrifuge tubes (1.5 ml, 2.0 ml)
- 4. Microcentrifuge capable of speed at ~18,000 rpm
- 5. 100 % ethanol
- Vortexer
- 7. Tube heater set to 60 °C and 70 °C for 1.5 or 2.0 ml microcentrifuge tubes

# **Working Buffer Preparations:**

#### ■ Preparation of VNE-Carrier RNA Buffer

Add 1 ml of VNE Buffer to the tube containing lyophilized Carrier RNA.Mix well by vortexing and transfer the mixture to the VNE Buffer when first open. Store the VNE-Carrier RNA Buffer at 4 °C.

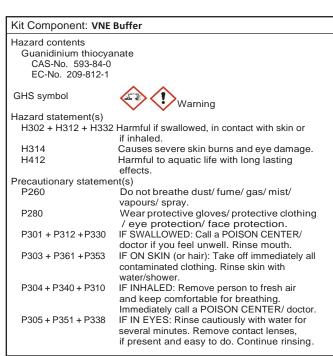
## □. Preparation of Wash Buffer 1 and Wash Buffer 2

Add required ethanol (96~100%) as the table below indicates. Store the Wash Buffer 1 & 2 (ethanol added) at  $15\sim25$  °C.

Cat. No:	FAVNK000-1 (4 preps)	FAVNK001 (50 preps)	FAVNK001-1 (100 preps)	FAVNK 001-2 (300 preps)
Ethanol for Wash Buffer 1	0.72 ml	18 ml	36 ml	108 ml
Ethanol for Wash Buffer 2	6 ml	80 ml	80 ml	200 ml

## **Safety Information:**

**CAUTION:** VNE Buffers and Wash Buffer 1 contain guanidinium salts which can form highly reactive compounds when combined withbleach. <u>DO NOT ADD BLEACH OR ACIDIC</u> SOLUTIONS DIRECTLY TO THE PREPARATION WASTE.



Hazardous conter	nts
	hloride, 20~50%, CAS-No. 50-01-1
Guariluine nyuroci	moriae, 20~3076, CA3-No. 30-01-1
GHS symbol	<b>!</b> Warning
Hazard statement(	s)
H302	Harmful if swallowed.
H319	Causes serious eye irritation.
Precautionary stat	tement(s)
P264	Wash thoroughly after handling.
P280	Wear protective gloves/ protective
	clothing/ eye protection/ face protection.

## **Important Notes:**

## 1. Notes for sample preparation:

- Make sure everything is RNase-free when handling this system.
- Buffers provided in this system contain irritants. Wear gloves, safety glasses, face protection, and a lab coat when handling these buffers.
- Do not thaw the frozen plasma or serum samples more than once.
- Centrifuge the plasma or serum samples at 6,000 x g for 3 minutes if precipitates are visible. Then transfer the cleared supernatant to a new vial and process itimmediately.

## 2. Notes for Buffers:

- •Add Carrier RNA to the VNE Buffer when first open. Store the VNE-Carrier RNA Buffer at 4 °C. see *Working Buffer Preparation*.
- Add required ethanol (96-100%) to Wash Buffer 1 and Wash Buffer 2 before use. see Working Buffer Preparation.
- For handling the buffers safely please read *Safety Information* before starting the procedure.

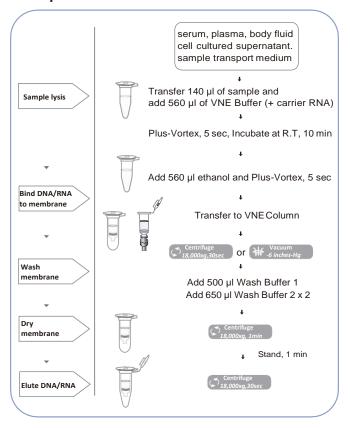
## 3. Notes for centrifugation and vacuuming:

- Ensure that centrifugation speed corresponds to the instruction of individual steps.
- When using the vacuum to proceed "DNA/RNA to column membrane" and "Wash column membrane", ensure that the tip of the column fits nicely into the manifold adaptor, and vacuum pressure has a capacity of achieving - 6 inches Hg.

• Units and values at same pressure (1 atm)

Office and values at same pressure (1 ati	11)
unit	value
atmosphere (atm)	1.000
millimeters of mercury (mmHg)	760.000
inches of mercury (inHg)	29.290
pascal (Pa)	101,325.000
kilopascal (KPa)	101.325
torr (torr)	760.000
pound per square inch (psi, 1bs/in²)	14.700

## **Brief procedure:**



1

## **Centrifugation Protocol:**

Please read Important Notes before continuing with these steps.

#### Sample type

## A. Cell-free fluid such as serum, plasma, body fluids and cellcultured supernatant

1-A1. Briefly spin the tube to descend the drops attached to the

Note! Centrifuge the sample at 7,000 x g for 3 minutes if precipitates are visible.

1-A2. Transfer 140 µl of the fluid sample (cleared supernatant) to a microcentrifuge tube (not provided).

#### B. Medium of transport swabs

- 1-B1. Briefly vortex the swab's transport tube, then briefly spin the tube to descend the drops attached to the tube wall.
- 1-B2. Transfer 140 µl of the medium to a microcentrifuge tube (not provided).

## • Sample lysis

2. Add 560 µl of VNE - Carrier RNA Buffer (Carrier RNA added, see Working Buffer Preparation). Mix well by vortexing and incubate for 10 minutes at room temperature.

## • Optimization of binding condition

3. Add 560 µl of ethanol (96~100 %) to the sample mixture, and mix well by plus-vortexing.

## • Bind viral DNA/RNA to column membrane (centrifuge)

- 4. Combine a VNE column with a Collection Tube (provided). Transfer up to 700 µl of sample mixture (ethanol added) to the VNE Column. Centrifuge at 8,000 x g for 1 min, then discard the flow-through. Combine the VNE Column with the used Collection Tube.
- 5. Transfer the rest of the sample mixture (ethanol added) to the VNE Column and centrifuge at 8,000 x g for 1 min. Discard the flow-through and the Collection Tube. Combine the VNE Column with a new Collection Tube (provided).

## • Wash column membrane (centrifuge)

- 6. Add 500 µl of Wash Buffer 1 (ethanol added) to the VNE Column. Centrifuge at 8,000 x g for 1 min then discard the flow-through. Combine the VNE Column with the used Collection Tube.
- Make sure that ethanol (96~100%) has been added into Wash Buffer 1 when first open.
- 7. Add 650 µl of Wash Buffer 2 (ethanol added) to the VNE Column. Centrifuge at 8,000 x g for 1 min then discard the flow-through. Combine the VNE Column with the used Collection Tube.
- Make sure that ethanol (96~100%) has been added into Wash Buffer 2 when firstopen.
- 8. Repeat step 7.

## Drv membrane

9. Centrifuge at full speed (~18,000 X g) for 1 min to dry the VNE Column. Discard the flow-through and the Collection Tube. ATTENTION: This step will avoid the residual liquid to inhibit the subsequent enzymatic reactions.

## Flute Viral RNA

- 10. Combine the VNE Column with an Elution Tube (provided). Add 40 ~ 60 µl of RNase-free Water to the membrane center of the VNE Column, Stand VNE Column for 1 min. ATTENTION: For effective elution, make sure that the RNase-free Water is dispensed onto the membrane center and is absorbed completely.
- 11. Centrifuge at full speed (~18.000 X a) for 1 min to elute the viral DNA/RNA. Store the viral DNA/RNA at -70 °C.

## Vacuum Protocol:

Please read Important Notes before continuing with these steps.

A. Cell-free fluid such as serums, plasma, body fluids and cellcultured supernatants

- 1-A1. Briefly spin the tube to descend the drops attached to the tube wall. Note! Centrifuge the sample at 7,000 x g for 3 minutes if the precipitates are visible.
- 1-A2. Transfer 140 µl of the fluid sample (cleared supernatant) to a microcentrifuge tube (not provided).

## B. Medium of transport swabs

- 1-B1. Briefly vortex the swab's transport tube, then briefly spin the tube to descend the drops attached to the tube wall.
- 1-B2. Transfer 140 µl of the medium to a microcentrifuge tube (not provided).

#### Sample lysis

2. Add 560 µl of VNE - Carrier RNA Buffer (Carrier RNA added, see Working Buffer Preparation). Mix well by vortexing and incubate for 10 minutes at room temperature.

## • Optimization of binding condition

3. Add 560 µl of ethanol (96~100 %) to the sample mixture and mix well by plus-vortexing.

## Bind viral DNA/RNA to column membrane (vacuum)

- 4. Combine the tip of a VNE Column with the adaptor of the vacuum manifold. Retain the Collection Tube to use on step 9. Transfer up to 700 µl of sample mixture (ethanol added) to the VNE Column and apply vacuum at -6 inches Hg until the column is empty. Switch off the vacuum and release the vacuum from the manifold.
- 5. Transfer the rest of the sample mixture (ethanol added) to the VNE Column, and apply vacuum at -6 inches Hg until the column is empty. Switch off the vacuum and release vacuum from the manifold.

## Wash column membrane (vacuum)

- 6. Add 500 µl of Wash Buffer 1 (ethanol added) to the VNE Column. Apply vacuum at -6 inches Hg until the column is empty. Switch off the vacuum and release the vacuum from the manifold.
- 7.Add 650 µl of Wash Buffer 2 (ethanol added) to the VNE Column, and apply vacuum at -6 inches Hg until the column is empty. Switch off the vacuum and release the vacuum from the manifold. 8. Repeat step 7.

## Drv membrane

9. Remove the VNE Column from the manifold and return the VNE Column back to the Collection Tube. Centrifuge at full speed (~18,000 X g) for 1 min to dry the VNE Column. Discard the flow -through and the Collection Tube.

Important step! This step will avoid the residual liquid to inhibit the subsequent enzymatic reactions.

## Elute Viral RNA

- 10. Combine the VNE Column with an Elution Tube (provided). Add 40 ~ 60 ul of RNase-free Water to the membrane center of the VNE Column. Stand VNE Column for 1 min. Important step! For effective elution, make sure that the RNase-free Water is dispensed onto the membrane center and is absorbed completely.
- 11. Centrifuge at full speed (~18.000 X g) for 1 min to elute the viral DNA/RNA. Store the viral DNA/RNA at -70 °C.

## **Troubleshooting**

## • Low yield

- Carrier RNA does not attach to VNE Buffer or VNE-Carrier Buffer is not stored well
- $\ \square$  Add 1 ml of VNE Buffer to Carrier RNA.Mix well; transfer it to the VNE Buffer, and store the VNE-Carrier RNA Buffer at 4 °C.
- Sample is not stored well or is thawn repeatedly
- □ Store samples at 80 °C for long-term storage. Frozen samples must not be thawed more than once.
- RNA Degradation
- ☐ Harvested samples are not immediately stabilized.
- Insufficient mixing with VNE-Carrier RNA Buffer
- ☐ Mix the sample mixture by plus-vortexing
- Insufficient lysis of protein
- □ Incubate the sample mixture at room temperature for 10 minutes after adding VNE-Carrier RNA Buffer.
- Improper RNA binding condition
- □ No ethanol added to the lysate (step 3) or incorrect percentage of ethanol was used.
- Incorrect RNA elution
- □ Ensure that RNase free water was added to the center of the VNE column membrane and was absorbed by the membrane.
- Incorrect preparation of Wash Buffer 1&2
- □ Ensure that the correct volume of ethanol (96~100 %) was added to Wash Buffers1&2 upon first use.

#### • Eluted RNA does not perform well

- Residual ethanol contamination
- ☐ Ensure that the VNE Column is centrifuged for 1 additional min at a speed of ~18,000 x g (step 9) after washing step.

## **FAVORGEN Products**:

For more product information, please visit our web site at www.favorgen.com

## Nucleic Acid Extraction - spin column (silica membrane)

- Viral DNA/ RNA Kit
- Viral Nucleic Acid Extraction Kit II
- Viral RNA/ DNA Vacuum Kit
- · Circulating Nuleic Acid Isolation Kit

## RNA Extraction - spin column (silica membrane)

- Blood/Cultured Cell Total RNA Mini/ Maxi Kit
- Soil RNA Isolation Mini Kit
- Tissue Total RNA Mini/Maxi Kit
- Plant Total RNA Mini/ Maxi Kit
- After Tri-Reagent RNA Clean-Up Kit

## 96-Well high throughput DNA/ RNA extraction (silica membrane)

- 96-well Gel/ PCR purification kit
- 96-well PCR Clean-Up Kit
- 96-Well Total RNA Kit
- 96 well Viral DNA/RNA extraction kit
- 96-Well Genomic DNA Extraction Kit
- 96-Well Plasmid Kit

#### Plasmid Extraction

- Mini/ Midi/ Maxi plasmid kit spin column (silica membrane)
- Midi/Maxi plasmid kit gravity flow column (anion
- Endotoxin Free Midi/Maxi plasmid kit gravity flow column (anion-exchange resin)

## DNA Clean-Up - spin column (silica membrane)

- PCR Clean-UP Kit/ GEL Purification Kit/ GEL/PCR Purification Kit
- MicroElute GEL/PCR Purification Kit

## DNA Extraction - spin column (silica membrane)

- Blood / Cultured Cell Genomic DNA Extraction Mini / Midi/
- Plant Genomic DNA Extraction Mini/ Maxi Kit
- Food DNA Extraction Kit
- · Milk Bacterial DNA Extraction Kit
- Tissue Genomic DNA Extraction Mini Kit
- FFPE Tissue DNA Extraction MicroElute Kit
- Fungi/ Yeast Genomic DNA Extraction Mini Kit
- Soil DNA Isolation Mini Kit • Stool DNA Isolation Mini Kit

## **Extraction Reagent**

• Tri-RNA Reagent - (Acid Guanidinium Thiocyanate-Phenol-Chloroform Extraction)

> Telephone: +886-8-762-1829 (Taiwan) Fax: +886-8-762-0791(Taiwan) web site: www.favorgen.com E-Mail: order@favorgen.com technical@favorgen.com

3